

CANNABINOID-BASED DRUGS AS ANTI-INFLAMMATORY THERAPEUTICS

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Abstract | In the nineteenth century, marijuana was prescribed by physicians for maladies ranging from eating disorders to rabies. However, as newer, more effective drugs were discovered and as the potential for abuse of marijuana was recognized, its use as a therapeutic became restricted, and only recently has its therapeutic potential been re-evaluated. Recent studies in animal models and in humans have produced promising results for the treatment of various disorders — such as obesity, cancer, and spasticity and tremor due to neuropathology — with drugs based on marijuana-derived cannabinoids. Moreover, as I discuss here, a wealth of information also indicates that these drugs have immunosuppressive and anti-inflammatory properties; therefore, on the basis of this mode of action, the therapeutic usefulness of these drugs in chronic inflammatory diseases is now being reassessed.

CANNABINOID RECEPTORS
G-protein-coupled receptors for Δ^9 -tetrahydrocannabinol, its synthetic analogues and endocannabinoids. They have been identified in most vertebrate phyla. Two subtypes are known: cannabinoid receptor 1 (CB₁) and CB₂.

ENDOCANNABINOID
Endogenous agonists for cannabinoid receptors that are present in animals. They are metabolites of eicosanoid fatty acids.

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Cannabis sativa — also known as marijuana — is the most frequently used illicit drug in the United States, particularly among young people, with up to 46.1% of those aged 17–18 being users. Although marijuana usage rates are lower than those for legal drugs, such as cigarettes and alcohol, they are significantly higher than those for other illicit drugs, such as cocaine and ecstasy. As a therapeutic, marijuana has been recognized for centuries, and in the nineteenth century, it was recommended as an analgesic, muscle relaxant, appetite stimulant and anticonvulsant¹. Its therapeutic applications continued to grow such that, by the early twentieth century, therapy with marijuana was used to ease the symptoms of a broad spectrum of diseases, ranging from rheumatism and epilepsy to tetanus and gonorrhoea. However, the popularity of marijuana as a therapeutic declined as newer, more effective drugs were discovered for these ailments and as the potential for abuse of marijuana was recognized, so by the early 1940s, it was removed from use as a pharmaceutical in the United States. From that time until the 1980s, marijuana and its derivatives, such as Δ^9 -tetrahydrocannabinol (THC), have been viewed as both scientific curiosities and illegal

drugs with a high potential for abuse; however, this now seems to be changing. In the past few years, numerous publications have reported the potential use of marijuana-based medicines for the treatment of diseases ranging from cancer to glaucoma^{1–8}, and one compound, SR141716A (also known as rimonabant or Acomplia; Sanofi-Synthélabo)⁹, has been widely publicized as the next wonder drug for promoting weight loss and smoking cessation. Here, I review the effects of marijuana and related compounds on the adaptive and innate immune systems and provide some perspective concerning the therapeutic potential of marijuana-derived cannabinoids and related compounds for the treatment of chronic inflammatory diseases.

Cannabinoid-based drugs and receptors

The main psychoactive component in marijuana is the ‘classical’ cannabinoid THC¹⁰ (FIG. 1a). Similar to its synthetic analogues (FIG. 1b,c), it functions by activating specific cell-surface CANNABINOID RECEPTORS, which are normally engaged by a family of endogenous ligands — the ENDOCANNABINOID (FIG. 1g,h). Compounds that bind these receptors induce CANNABIMIMETIC responses

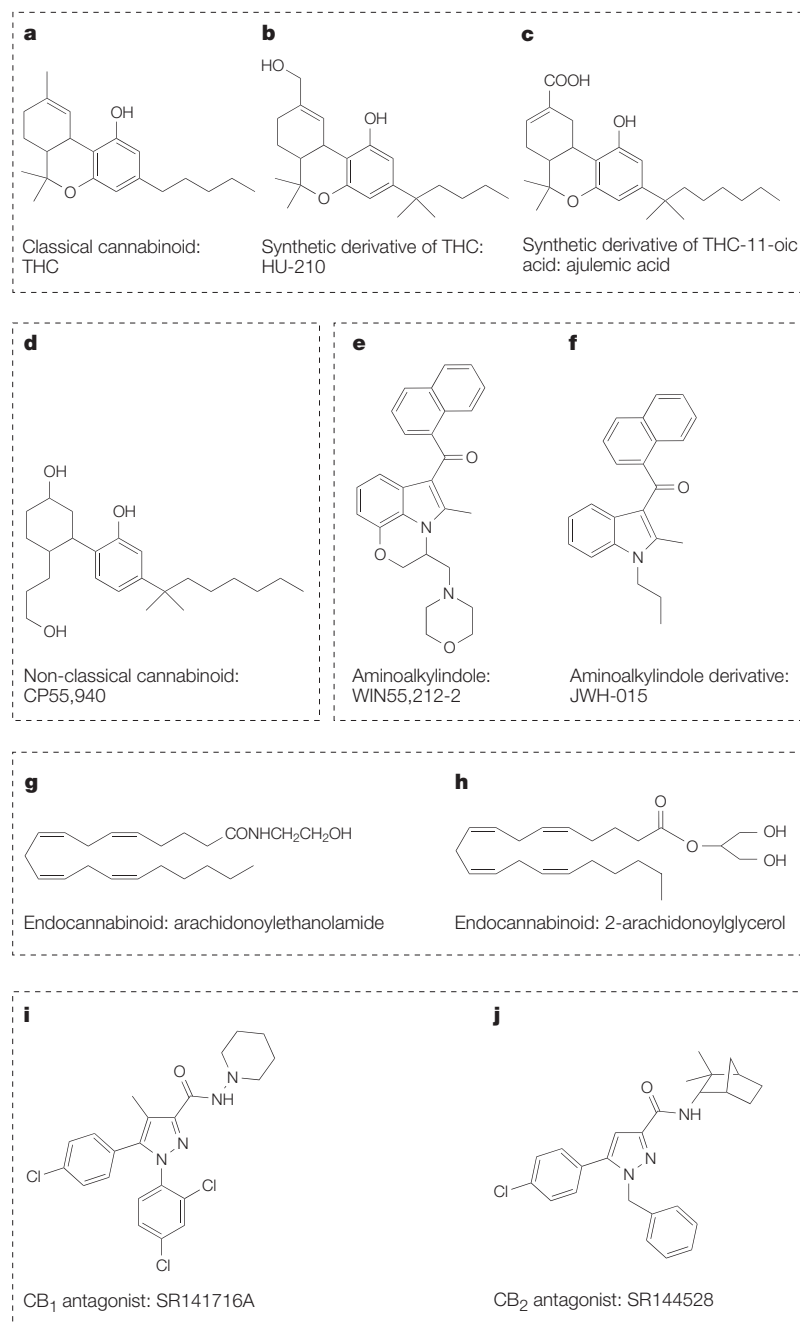


Figure 1 | Drugs based on marijuana-derived cannabinoids are divided into various groups. There are four groups of cannabinoids or cannabimimetic drugs that have cannabinoid-receptor-binding activity. The first group contains the ‘classical’ cannabinoids, which are tricyclic dibenzopyran derivatives. They occur naturally in the marijuana plant — for example, Δ^9 -tetrahydrocannabinol (THC) (**a**). Also, there are synthetic analogues of these natural compounds — for example, HU-210 (**b**). These cannabinoids bind relatively non-selectively to both cannabinoid receptor 1 (CB₁) and CB₂, with HU-210 having a higher affinity than THC. Ajulemic acid (**c**) is another analogue; it is a classical cannabinoid derived from THC-11-oic acid. Ajulemic acid binds both cannabinoid receptors but has a lower affinity for CB₂. The second group contains the non-classical cannabinoids, which are synthesized analogues that lack the dihydropyran ring of THC. This group contains compounds such as CP55,940 (**d**). The third group contains the aminoalkylindoles, such as WIN55,212-2 (**e**). Derivatives of WIN55,212-2, such as JWH-015 (**f**), have been shown to selectively bind CB₂. The fourth group contains the endocannabinoids, which are eicosanoid compounds rather than cannabinoid compounds. The most studied members of this group are arachidonylethanolamide (**g**) and 2-arachidonoylglycerol (**h**). Finally, several cannabinoid-receptor antagonists have been synthesized, and the most widely studied are SR141716A (**i**), which selectively binds CB₁, and SR144528 (**j**), which selectively binds CB₂.

in vivo and *in vitro*¹¹. Research on the structure–activity relationships of THC led to the synthesis of ‘non-classical’ cannabinoid analogues. The most widely studied of this group is CP55,940 (FIG. 1d), a high-affinity ligand with potent biological effects that are mediated through binding both cannabinoid receptor 1 (CB₁) and CB₂. Another class of cannabimimetic agents is the aminoalkylindoles, such as WIN55,212-2 (FIG. 1e), which also has high affinity for both receptors and has potent activity. By contrast, some derivatives of this class, such as JWH-015 (FIG. 1f), have been shown to selectively bind CB₂. Such derivatives might be of therapeutic value, because psychoactive effects are mediated through binding CB₁ and not through binding CB₂. Other plant-derived cannabinoids (such as cannabidiol) and synthetic derivatives (such as HU-211; also known as dexanabinol; Pharmos Corporation) bind CB₁ and CB₂ with very low affinity, so they have low cannabimimetic activity and might function by binding different receptors, such as the NMDA (N-METHYL-D-ASPARTATE) RECEPTOR⁵ or other unknown receptors. Furthermore, a derivative of THC-11-oic acid, ajulemic acid (also known as CT-3) (FIG. 1c), has low affinity for CB₂ (REF. 12) but has anti-inflammatory activity¹³, which might be mediated through disruption of the arachidonic-acid cascade¹⁴ or through activation of peroxisome-proliferative-activated receptor- γ (PPAR- γ)¹⁵. Finally, receptor antagonists have been synthesized, such as SR141716A and SR144528 (REF. 11) (FIG. 1i,j), which inhibit or reverse the biological effects of CB₁ and CB₂ agonists. These antagonists have been used experimentally to determine the receptor-binding activities of various putative agonists and are now also being used clinically to inhibit CB₁ activity. For example, SR141716A is given to suppress appetite during the treatment of obesity⁹.

The naturally occurring endocannabinoids are produced by the cleavage of membrane fatty acids^{16,17}, in particular arachidonic acid, and have varying specificities for the two cannabinoid receptors. Arachidonylethanolamide (AEA; previously known as anandamide) is an endogenous fatty-acid amide and was the first endocannabinoid to be discovered¹⁸ (FIG. 1g). It has less intrinsic activity when bound to CB₂ than to CB₁, and it has also been shown to bind VANILLOID RECEPTORS, which are ligand-gated cation channels that are sensitive to capsaicin and related analogues¹⁹. Several other endocannabinoids have also been described, including 2-arachidonoylglycerol (2-AG)²⁰ (FIG. 1h) and 2-arachidonoylglyceryl ether (also known as noladin ether)²¹, with the former being a full agonist of CB₂ and the latter showing a higher affinity for CB₁. The endocannabinoids are produced by various cells, including cells of the immune system and the brain (discussed later).

In addition to the two cannabinoid receptors that have been characterized^{22,23}, there might be others, particularly receptors that interact with the endocannabinoids, which would be consistent with the observation that AEA also binds vanilloid receptors¹⁹. CB₁ and CB₂ are seven-transmembrane, G-protein-coupled receptors that are coupled to G_i or G_o heterotrimeric proteins and adenylyl cyclases, but other second messengers and

CANNABIMIMETIC
 Δ^9 -Tetrahydrocannabinol (THC)-like in pharmacological terms. A compound is usually accepted to be cannabimimetic if it produces four characteristic effects of THC in an *in vivo* assay known as the 'mouse tetrad model'. These effects are hypomotility, hypothermia, analgesia and a sustained immobility of posture (catalepsy).

signalling components are also involved in their activity^{24,25}. The tissue distribution of CB₁ and CB₂ accounts for the well-known psychotropic and peripheral effects of cannabinoids (reviewed in REFS 11,25–28). CB₁ is abundant in the central nervous system (CNS), in particular in the basal ganglia, cerebellum, hippocampus and cortex, where the receptors and endocannabinoids are implicated in retrograde neurotransmitter regulation of synaptic transmission^{28,29}. CB₁ is also expressed in the periphery, as is CB₂, where they are mainly restricted to immune cells and tissues (discussed later).

Endocannabinoids in innate immunity

Production of endocannabinoids. Microbial pathogens that invade the tissues are recognized by host cells and host factors that trigger the activation of both innate and adaptive immune responses³⁰. Activation of the inflammatory response to infection largely depends on the release of pro-inflammatory cytokines and chemokines. However, in addition to cytokines and other proteins, various metabolic products of immune cells — including membrane fatty acids, such as arachidonic acid³¹ — have also been implicated in the inflammatory response to infection. It is therefore not surprising that

chemically similar metabolites — such as the endocannabinoid AEA — are produced and released by activated immune cells³² (FIG. 2). *In vitro* studies have shown that stimulation with bacterial lipopolysaccharide (LPS) increases the production of AEA and 2-AG by immune cells (FIG. 2), including macrophages³³, peripheral-blood mononuclear cells (PBMCs)³⁴ and dendritic cells³⁵. In addition, LPS-activated PBMCs show reduced expression of the AEA-degrading enzyme fatty-acid amide hydrolase (FAAH)³⁶. Given that inflammatory responses must be tightly controlled to avoid extensive tissue damage³⁷ and to allow a return to homeostatic conditions, FAAH-mediated degradation is one of the mechanisms for the inactivation of endocannabinoids. In addition, as-yet-uncharacterized endocannabinoid membrane transporters seem to facilitate both the release and the subsequent uptake of endocannabinoids by neurons and glial cells, thereby contributing to the regulation of endocannabinoids. So, immune cells from humans and animals increase the production of endocannabinoids in response to LPS and in response to activation by other stimuli. Additional studies are needed to determine the range of immune and microbial stimuli that induce endocannabinoid production and to further define the mechanisms that regulate this effect.

Endocannabinoids as chemotactic agents. After recognition of an invading pathogen, the release of cytokines and chemokines by cells involved in the innate immune response triggers an influx of lymphoid and myeloid cells from the blood to the site of infection³⁸. Recent evidence indicates that the endocannabinoid 2-AG also induces the migration of various cell types (FIG. 2). For example, in transwell cultures, 2-AG attracted human eosinophils³⁹, as well as Raji B cells⁴⁰ and mouse bone-marrow-derived dendritic cells⁴¹. Furthermore, CB₂ seems to be overexpressed in several myeloid leukaemias, and these leukaemic cells are induced to migrate after stimulation with 2-AG⁴². It therefore seems that endocannabinoids might be involved in cellular migration by functioning as chemotactic agents, together with cytokines and proteins that have classically been defined as chemokines. It is interesting to note that a similar function has been reported for opioids, such as morphine, which have neuroimmune functions in common with cannabinoids⁴³.

Expression of cannabinoid receptors. In addition to the induction of endocannabinoid expression and the subsequent chemotaxis of immune cells, there is evidence that activation of immune cells by LPS or other stimuli also modulates the expression of the cannabinoid receptors CB₁ and CB₂ by these cells (FIG. 2). For example, the initial report on the cloning of CB₂ also showed that stimulation of the human pro-myelocytic leukaemia cell line HL-60 with phorbol 12-myristate 13-acetate (PMA) caused an increase in the level of mRNA encoding CB₂ (REF 23); similar results were also obtained in mouse splenocyte cultures stimulated with CD40-specific antibody⁴⁴. In addition to CB₂, the levels of mRNA encoding CB₁ were also increased in

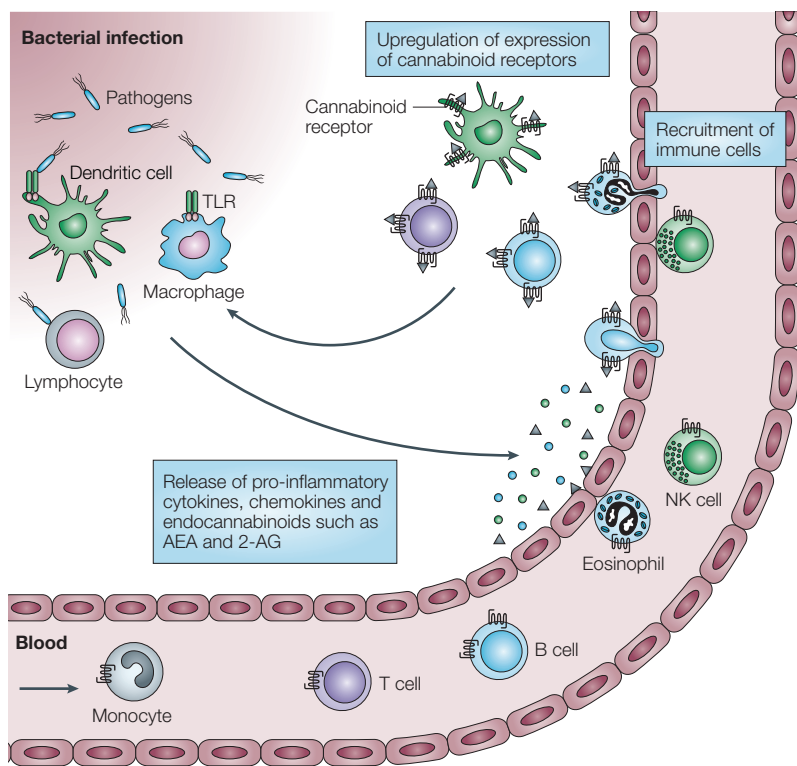


Figure 2 | **The endocannabinoid system and innate immunity.** Bacteria stimulate lymphocytes, dendritic cells (DCs) and macrophages — through pattern-recognition receptors, such as Toll-like receptors (TLRs) — to release cytokines and chemokines, which attract leukocytes to the site of infection. It is now known that stimulation of these cells also induces the release of endocannabinoids — such as arachidonylethanolamide (AEA) and 2-arachidonoylglycerol (2-AG) — that are also chemotactic for leukocytes. Leukocytes — including T cells, B cells, eosinophils, natural killer (NK) cells, DCs and macrophages — invade the tissues, promoting the elimination of the microorganisms and the development of an adaptive immune response. These activated cells seem to upregulate the expression of both types of cannabinoid receptor — cannabinoid receptor 1 (CB₁) and CB₂ — and these are then available for participation in immune regulation.

both Jurkat T cells⁴⁵ and mouse splenocytes⁴⁶ after stimulation. However, other studies have shown that stimulation decreased the expression of these receptors^{46,47}, which most probably reflects differences in the stimulatory substance used or the cell type studied. LPS has also been shown to modulate cannabinoid-receptor expression, although findings from separate studies are inconsistent, with some reports showing upregulation of receptor expression⁴⁸ and others showing downregulation^{35,44,49}.

Consistent with a possible role for endocannabinoids and CB₂ in chemotaxis and other immune-activation events, a recent study of mouse brain microglial cells showed that, under chemotactic conditions, the expression of CB₂ was specifically localized to the leading edge of the cell⁵⁰, indicating that there is receptor modulation similar to that observed for immunoreceptors during chemotaxis or during the formation of the immunological synapse⁵¹. Taken together, these studies indicate that microbial antigens or other stimuli that induce immune activation influence cannabinoid-receptor expression by immune cells; however, the factors that are involved in increased or decreased expression are far from understood, and little is known about the molecular regulation of the genes encoding these receptors in immune cells. Further research is needed to take full advantage of manipulating this potentially important immunomodulating system for therapeutic purposes (discussed later).

Cannabinoids in inflammation

Cannabinoids modulate cytokine production. In the mid-1980s, it was shown that mouse cells treated with the drug THC produced decreased levels of type I interferons (IFN- α and IFN- β) after stimulation with LPS or polyinosinic-polycytidylic acid (polyI:C)^{52,53}, providing the first evidence that cannabinoids might modulate cytokine production. Many subsequent studies have shown that cannabinoids, for the most part, suppress the production of cytokines in innate and adaptive immune responses, both in animal models and in human cell cultures^{26,27,54}. Their suppression of pro-inflammatory cytokine and chemokine production indicates that these drugs might have anti-inflammatory effects and could therefore be used for the treatment of chronic inflammatory diseases. Consistent with this, serum levels of tumour-necrosis factor (TNF) and interleukin-12 (IL-12) were shown to be decreased in mice that were primed by infection with *Propionibacterium acnes* (*Corynebacterium parvum*) and stimulated with an injection of LPS (conditions that promote optimum cytokine upregulation) then treated with the synthetic THC derivative HU-210 or the aminoalkylindole WIN55,212-2 (REF. 55). The cannabinoids also protected these mice from the lethal effects of LPS in this model, and this protection might have resulted from, at least in part, a concomitant drug-induced increase in the levels of the regulatory cytokine IL-10 (TABLE 1). TNF production was also suppressed in the brain of rats subjected to closed head injury, after treatment with the non-psychoactive cannabinoid HU-211, and

this treatment was also neuroprotective and resulted in a better clinical outcome⁵⁶. Although HU-211 does not bind cannabinoid receptors and is therefore not cannabimimetic, it seems to function as an **NMDA-receptor** antagonist, thereby preventing excitotoxicity and neuronal death⁵⁷. In another mechanistically complex mouse model, treatment with WIN55,212-2 decreased tissue damage after myocardial ischaemia-reperfusion injury⁵⁸. Treatment of animals before ischaemia and reperfusion considerably reduced the size of the infarct, and this was paralleled by lower levels of production of IL-1 β and CXC-chemokine ligand 8 (CXCL8) in the injured tissue. In human studies, lung alveolar macrophages removed from marijuana smokers were compromised in their ability to produce TNF, granulocyte/macrophage colony-stimulating factor and IL-6 in response to LPS stimulation⁵⁹ (TABLE 1). It therefore seems that cannabinoids can inhibit the production of TNF and other cytokines in several different models and by several different mechanisms (TABLE 1), not all of which depend on interaction with cannabinoid receptors.

As well as suppressing the production of cytokines, cannabinoids have been shown to increase the production of cytokines (including TNF, IL-1, IL-6 and IL-10) when they are administered together with bacteria or other antigens^{60–62}, or in some cases, when cannabinoids are administered alone^{63,64}. So, *in vivo*, cannabinoids might either suppress or enhance the production of these pro-inflammatory agents, depending on either the nature of the pro-inflammatory stimulus or the type of cannabinoid used (TABLE 1).

Cannabinoids modulate inflammatory-cell migration.

The effect of cannabinoids has also been analysed in the experimental allergic encephalomyelitis (EAE) model of **multiple sclerosis** (a neuroinflammatory disease), and they have been shown to suppress disease progression^{65,66} and inflammatory reactions⁶⁶. In one study, mice were induced to develop EAE and given WIN55,212-2 alone or WIN55,212-2 and then either a CB₁ or CB₂ antagonist every 4 days. After several weeks, disease progression was assessed, together with leukocyte-endothelial-cell interactions, using intravital microscopy. Treatment with WIN55,212-2 suppressed the rolling and adhesion of venous leukocytes and improved neurological function in mice with EAE, compared with control animals; furthermore, the suppressive effect was attenuated by co-treatment with a CB₂ antagonist but not a CB₁ antagonist, indicating that CB₂ is involved in these processes⁶⁶. The reduction in **DIAPYDESIDIS** presents a paradox, because cannabinoids have been shown to enhance chemotaxis and chemokine production (as discussed earlier). Although the levels of cytokines and adhesion molecules were not measured in this study, it is possible that cannabinoid-mediated suppression of leukocyte adhesion results from an inhibition of T helper 1 (T_H1)-cell cytokine production: for example, of IFN- γ , which facilitates transendothelial cell trafficking^{67,68} (discussed later).

NMDA RECEPTOR

(*N*-methyl-D-aspartate receptor). NMDA is a synthetic amino acid with affinity for NMDA receptors, which mediate excitatory effects in the brain when they are stimulated by endogenous ligands such as glutamic acid. Overstimulation can lead to neuronal excitotoxicity.

VANILLOID RECEPTORS

Cation channels that are expressed by nerve sensory fibres and are involved in the perception of pain. These receptors are ligand-, proton- and heat-activated and are targets for capsaicin — the hot component of chillies.

DIAPYDESIDIS

The last step in the leukocyte-endothelial-cell adhesion cascade. This cascade includes tethering, triggering, tight adhesion and transmigration. Diapedesis is the migration of leukocytes across the endothelium, which occurs by squeezing through the junctions between adjacent endothelial cells.

Table 1 | **Cannabinoid effects on pro-inflammatory cytokines**

Cannabinoid	Receptor	Cell or tissue type	Cytokine stimulant or inflammation model	Effect	Reference
Mice					
WIN55,212-2 or HU-210	ND	Spleen	LPS and <i>Propionibacterium acnes</i> *	Decreases TNF and IL-12 and increases IL-10	55
THC	ND	Macrophage cell line (RAW264.7)	LPS	Decreases TNF	122
HU-211	NMDA receptor	Brain	Closed head injury	Decreases TNF	56
WIN55,212-2	CB ₂ dependent	Heart	Ischaemia-reperfusion	Decreases IL-1β and CXCL8	58
THC, AEA or 2-AG	ND	Macrophage cell line (J774)	LPS	Decreases IL-6	123
THC	ND	Spleen	<i>Legionella pneumophila</i>	Increases TNF and IL-6	60
THC	ND	Peritoneal macrophages	LPS	Increases IL-1α and IL-1β	61
Humans					
Marijuana smoking	ND	Lung alveolar macrophages	LPS	Decreases TNF, GM-CSF and IL-6	59
Ajulemic acid	ND	Peripheral-blood and synovial monocytes	LPS	Decreases IL-1β	100
2-AG	CB ₂ dependent	Pro-myelocytic leukaemia cell line (HL-60)	2-AG	Increases CXCL8 and CCL2	64
CP55,940	CB ₂ dependent	Pro-myelocytic leukaemia cell line (HL-60) (CB ₂ transfected)	CP55,940	Increases TNF, CXCL8, CCL2 and CCL4	63
Rats					
AEA, 2-AG, WIN55,212-2 or HU-210	CB ₁ and CB ₂ independent	Microglial cells	LPS	Decreases TNF	124

**Propionibacterium acnes* was previously known as *Corynebacterium parvum*. AEA, arachidonylethanolamide; 2-AG, 2-arachidonoylglycerol; CB, cannabinoid receptor; CCL, CC-chemokine ligand; CXCL8, CXC-chemokine ligand 8; GM-CSF, granulocyte/macrophage colony-stimulating factor; IL, interleukin; LPS, lipopolysaccharide; ND, not determined; NMDA, *N*-methyl-D-aspartate; THC, Δ⁹-tetrahydrocannabinol; TNF, tumour-necrosis factor.

An increasing number of studies indicate that cannabinoids and related compounds modulate pro-inflammatory cytokines in various systems, ranging from infection to tissue injury. So far, the mechanisms of this modulation are unclear, but mechanisms that involve cannabinoid receptors, as well as other mechanisms, seem to be involved. For therapeutic applications, compounds that function either by binding CB₂ or by by-passing cannabinoid receptors altogether would be of benefit, because CB₁-mediated psychoactive side-effects would be diminished.

Cannabinoids modulate T helper cells

Cannabinoid treatment has been shown to suppress both innate immunity and adaptive immunity, and the effects of cannabinoids on humoral and cellular immunity have been extensively reviewed^{26,27,69}. So, here I focus on the more recent studies that indicate that cannabinoids have a T_H-cell biasing effect, in which T_H1-cell activity is suppressed and T_H2-cell activity is increased (TABLE 2). The first indications of this effect can be traced to the observation that treatment with THC induces suppression of both T_H1-cell activity and cell-mediated immunity in mice infected with *Legionella pneumophila*⁷⁰. Additional studies showed that treatment of mice with THC not only decreased the production of IFN-γ and the levels of IL-12 and the IL-12 receptor (IL-12R) but also increased the production of IL-4 (REF. 71) (TABLE 2), showing that the immune

response to the drug was biased towards the production of T_H2 cells. Cannabinoid receptors were shown to be involved in the effects of THC, and it is possible that signalling through these G-protein-coupled receptors suppresses the expression of IL-12, as has been shown in other systems⁷². In addition to this infection model, the T_H-cell-biasing effect was also shown in a mouse model of lung cancer, in which IFN-γ production was decreased but IL-10 and transforming growth factor-β (TGF-β) production were increased⁷³. The presence of higher levels of these cytokines indicates that THC either biases the immune response towards T_H2 cells or activates regulatory T cells, either T regulatory 1 (T_R1) cells or T_H3 cells^{74,75}. These regulatory T-cell populations have been shown to produce IL-10 and TGF-β, respectively, so they could be the source of these cytokines following treatment with THC. THC was also shown to bias T_H-cell differentiation towards T_H2 cells during an allogeneic response to human peripheral-blood T cells⁷⁶. Furthermore, PBMCs isolated from marijuana smokers proliferated less and produced less IL-2 in response to mitogens, but they produced more IL-10 and TGF-β than PBMCs isolated from non-marijuana smokers, indicating a T_H-cell bias towards T_H2 cells⁷⁷ (TABLE 2). Animal models of multiple sclerosis have also shown that treatment with cannabinoids attenuates cell-mediated immunity and T_H-cell activity, concomitant with reducing disease symptoms. For example, in a model in which demyelinating disease is induced by Theiler's murine

encephalomyelitis virus, cannabinoid-receptor ligands — such as WIN55,212-2 and JWH-015 — were shown to ameliorate disease progression by reducing the delayed-type hypersensitivity response, by decreasing IFN- γ production⁷⁸ and microglial-cell activation, and by suppressing the number of CD4⁺ T cells infiltrating the brain⁷⁹.

From these studies, it seems that cannabinoids bias the immune response away from T_H1-cell immunity and that cannabinoid receptors are involved in this process. It is possible that signalling through cannabinoid receptors expressed by T and B cells, as well as by antigen-presenting cells, suppresses the expression of T_H1-cell-promoting cytokines and increases the expression of T_H2-cell-promoting cytokines. Indeed, treatment with cannabinoids has been shown to alter the expression of specific transcription factors: the expression of the T_H2-cell-promoting transcription factor GATA-binding protein 3 (GATA3) has been found to be increased by treatment with THC^{80,81}, whereas the production of IL-2-expression-promoting transcription factors is suppressed by treatment with cannabinoids⁸². The selective suppression of T_H1-cell immunity by these drugs supports their potential use in the treatment of chronic inflammatory diseases.

Therapeutics for inflammatory diseases

Recent work has led researchers to consider that cannabinoid-based drugs have therapeutic potential for the treatment of a variety of disorders. For example, the CB₁ antagonist SR141716A has been shown to effectively promote weight loss and smoking cessation in preclinical and clinical trials^{9,83,84}. Furthermore, cannabinoids seem to be neuroprotective in models of inflammatory neurodegenerative disease, possibly because some derivatives function as inhibitors of NMDA receptors and as antioxidants^{5,7} and because they function to control spasticity and tremor that result from neurodegeneration⁸⁵. These drugs have also been proposed for the treatment of tumours, owing to their antiproliferative and pro-apoptotic effects^{2,86–88}. However, as cannabinoids can also modulate innate and adaptive immune

responses, their therapeutic potential is now being evaluated on the basis of their immunomodulatory and anti-inflammatory actions.

Nervous-tissue inflammation. Injection of THC was initially shown to suppress the neurological signs and symptoms of EAE in rat and guinea-pig models⁸⁹ (TABLE 3). A marked reduction in inflammation in the CNS of cannabinoid-treated animals was observed, and this was later suggested to result from a THC-induced increase in the serum levels of the steroid corticosterone, which is known to be anti-inflammatory and immunosuppressive⁹⁰. However, this explanation might be an oversimplification, because, as described earlier, cannabinoids attenuate demyelinating disease in these models by suppressing the differentiation of T_H cells into T_H1 cells^{78,79}, and this suppression can be uncoupled from THC-induced steroid mobilization in treated animals⁹¹. Another consequence of neurodegeneration is neuropathic or central pain, which results from sclerotic plaques that affect pain pathways in the CNS⁹². These lesions and central pain are observed in various chronic conditions, including trauma, type 2 diabetes and multiple sclerosis. Although cannabinoids have a well-recognized analgesic effect that is mediated by signalling through CB₁ (REF. 93), compounds that selectively bind CB₂ and act in the periphery have recently been shown to suppress experimentally induced central pain⁹⁴. The aminoalkylindole derivative AM1241 was shown to reverse tactile and thermal hypersensitivity produced in rats by spinal nerve ligation, and this effect was blocked by a CB₂ antagonist, but not a CB₁ antagonist, and occurred in CB₁-deficient mice. This almost completely CB₂-dependent effect has been proposed to occur in the periphery because of the paucity of these receptors in the CNS; the mechanism of action has been suggested to be indirect, with the drug acting on local mast cells and immune cells to suppress the release of mediators (such as histamines and TNF) that can sensitize primary afferent neurons, thereby rendering them more sensitive to pain⁹⁴. This anti-inflammatory mechanism has yet to be established; however, it is

Table 2 | Cannabinoid effects on adaptive immunity and T helper cells

Cannabinoid	Receptor	Cell or tissue type	Cytokine stimulant or inflammation model	Effect	Reference
Mice					
THC	ND	Spleen	<i>Legionella pneumophila</i>	Decreases IFN- γ and IgG2a	70
THC	CB ₁ and CB ₂ dependent	Spleen	<i>Legionella pneumophila</i>	Decreases IL-12 and IL-12R and increases IL-4	71
THC	CB ₂ dependent	Spleen	Tumour model	Decreases IFN- γ and increases IL-10 and TGF- β	73
WIN55,212-2	ND	Spleen	Theiler's murine encephalomyelitis virus	Decreases IFN- γ	78
Humans					
THC	CB ₂ dependent	Peripheral-blood T cells	Allogeneic dendritic cells	Decreases IFN- γ	76
Marijuana smoking	ND	Peripheral-blood mononuclear cells	Phytohaemagglutinin and concanavalin A	Decreases IL-2 and increases IL-10 and TGF- β	77

CB, cannabinoid receptor; IFN- γ , interferon- γ ; IL, interleukin; IL-12R, IL-12 receptor; ND, not determined; TGF- β , transforming growth factor- β ; THC, Δ^9 -tetrahydrocannabinol.

interesting that, in a recent small clinical trial, the THC-based drug dronabinol (Marinol; Unimed Pharmaceuticals, Inc.) was shown to have a small but clinically relevant analgesic effect on central pain in patients with multiple sclerosis; this is possibly associated with, in part, suppression of release of these sensitizing mediators through a CB₂-dependent mechanism⁹⁵. From these findings and other data, it seems that cannabinoids have several mechanisms of action for the attenuation of

symptoms and disease progression of multiple sclerosis and other neurodegenerative diseases. These mechanisms might include the inhibition of T_H1-cell responses in the CNS, owing to increased steroid production and other mechanisms, and the suppression of neuro-immune mediators of neuropathic pain that are locally released (FIG. 3). In addition to these mechanisms, the action of cannabinoids in reducing spasticity, which does not seem to be mediated by inflammatory

Table 3 | **Preclinical and clinical studies examining the anti-inflammatory effects of cannabinoids**

Preclinical model	Drugs	Outcome or mechanism	Clinical trials	Drug	Outcome	Refs
Nervous-tissue inflammation						
EAE in rats and guinea pigs	THC	Decreased CNS inflammation	–	–	–	89
EAE in rats	THC	Reduced disease progression and increased corticosterone production	–	–	–	90
Theiler's murine encephalomyelitis-virus-induced EAE in mice	WIN55,212-2	Decreased DTH responses and IFN- γ production	–	–	–	78
Theiler's murine-encephalomyelitis-virus-induced EAE in mice	WIN55,212-2, ACEA or JWH-015	Decreased CD4 ⁺ T-cell influx to CNS	–	–	–	79
Neuropathic pain in rats	AM1241	Decreased nerve hypersensitivity (CB ₂ involved) and pro-inflammatory-mediator production	Central pain in patients with multiple sclerosis	Dronabinol	Analgesia	94,95
Closed head injury in mice	2-AG	Decreased brain oedema and improved clinical outcome (CB ₁ involved)	Cerebral pressure and clinical outcome in patients with head injury; Phase III trial for patients with traumatic brain injury	HU-211 HU-211	Favourable No efficacy	100,101 102
Closed head injury in rats	HU-211	Decreased TNF production and neuropathology	–	–	–	56
Inflammatory bowel disease						
LPS-induced gastrointestinal transit in rats	ACEA or JWH-133	Decreased gastrointestinal transit (CB ₂ involved) and pro-inflammatory-mediator production	–	–	–	107
Chemically induced colitis in mice	HU-210	Decreased colonic inflammation (CB ₁ involved) and inflammatory-cell influx	–	–	–	108
Arthritis						
Leukocyte influx in mice and adjuvant-induced arthritis in rats	Ajulemic acid	Decreased granulocyte influx, joint inflammation and prostaglandin production	–	–	–	13
PBMCs and synovial-fluid monocytes	Ajulemic acid	Decreased IL-1 β production	–	–	–	110
Collagen-induced arthritis in mice	Cannabidiol	Decreased arthritis, cell-mediated immunity, and IFN- γ and TNF production	–	–	–	112
Collagen-induced arthritis in mice	HU-320	Decreased arthritis, cell-mediated immunity and TNF production	–	–	–	114
Vascular inflammation						
LPS-induced hypotension in rats	SR141716A	Decreased hypotension (2-AG and CB ₁ mediate hypotension)	–	–	–	115
Myocardial ischaemia-reperfusion injury in mice	WIN55,212-2	Decreased tissue injury and cytokine production (CB ₂ involved)	–	–	–	58
Septic shock in mice	HU-211	Decreased lethality and TNF production	–	–	–	117

ACEA, arachidonoyl-2'-chloroethylamide; 2-AG, 2-arachidonoylglycerol; CB, cannabinoid receptor; CNS, central nervous system; DTH, delayed-type hypersensitivity; EAE, experimental allergic encephalomyelitis; IFN- γ , interferon- γ ; IL-1 β , interleukin-1 β ; LPS, lipopolysaccharide; PBMC, peripheral-blood mononuclear cell; THC, Δ^9 -tetrahydrocannabinol; TNF, tumour-necrosis factor.

changes⁹⁶, has been analysed in clinical trials and has been shown to have efficacy in reducing spasms and stiffness^{97–99}.

Several studies have shown that endocannabinoids (such as AEA and 2-AG), as well as synthetic cannabinoids that bind cannabinoid receptors with very low affinity (such as HU-211), are neuroprotective after traumatic brain injury. The mechanisms that mediate the protective effects involve cannabinoid-receptor function, as well as NMDA receptors, inhibition of GLUTAMATERGIC SYNAPTIC TRANSMISSION, and reduction of TNF production and oxidative stress⁵. For example, levels of endogenous 2-AG were found to be markedly increased in mice with closed head injury, and administration of 2-AG following injury reduced brain oedema (a consequence of inflammation) and led to a better clinical outcome¹⁰⁰ (TABLE 3). The effect of 2-AG was attenuated by pretreatment with a CB₁ antagonist, indicating the involvement of cannabinoid receptors. However, receptor-independent mechanisms also seem to be involved. For example, administration of HU-211 attenuated neuropathology in rats with closed head injury and decreased TNF production in the brain, indicating that the suppression of cytokine production by HU-211 was at least partly responsible for the neuroprotection⁵⁶. Similarly, administration of HU-211 to patients with severe closed head injury was initially shown to result in a better clinical outcome¹⁰¹ but was recently reported not to have any clinical efficacy¹⁰². As well as TNF, other pro-inflammatory mediators, such as the IL-1R antagonist, might also be involved in the neuroprotective effects of cannabinoids¹⁰³. So, it seems that cannabinoids and endocannabinoids regulate some of the inflammatory aspects of brain injury and that this occurs by both cannabinoid-receptor-mediated and non-cannabinoid-receptor-mediated mechanisms. It is possible that these drugs reduce brain oedema and other aspects of neuroinflammation by inhibiting NMDA receptors, by functioning as antioxidants and by reducing the levels of pro-inflammatory cytokines in the brain (FIG. 3).

Inflammatory bowel disease. Inflammatory bowel disease, which includes ulcerative colitis and Crohn's disease, affects millions of individuals. Better therapies are needed to control the chronic inflammation that is associated with this disease, which leads to increased intestinal motility and faecal transit, resulting in pain, diarrhoea and poor ability to digest food³. The endocannabinoid system seems to be involved in the inhibition of intestinal motility, because functional CB₁ is expressed in the human ileum and colon¹⁰⁴, and its expression is increased during inflammation¹⁰⁵. Furthermore, AEA and 2-AG can be present in high levels in the gut¹⁰⁵ and can inhibit colonic propulsion in mice¹⁰⁶. Also, activation of CB₂ was recently shown to attenuate LPS-induced increases in gastrointestinal transit¹⁰⁷ (TABLE 3), and CB₁ agonists were shown to be suppressive — but only for basal, and not for LPS-induced, gastrointestinal transit. Interestingly, CB₂-agonist-mediated inhibition of LPS-induced

gastrointestinal transit was associated with the attenuation of inflammatory changes, such as suppressing the levels of cyclooxygenases and inducible nitric-oxide synthase, indicating that prostaglandins and/or nitric oxide mediate the effects of CB₂ agonists on gastrointestinal transit¹⁰⁷. Anti-inflammatory effects of cannabinoids were also observed in a mouse model of chemically induced colitis¹⁰⁸. Both colonic-tissue ulceration and the inflammatory response were attenuated by cannabinoid treatment, as well as in FAAH-deficient mice, as assessed histologically and by measuring tissue myeloperoxidase activity (which correlates with neutrophil influx). By contrast, ulceration and inflammation were increased in mice deficient in CB₁ and in mice treated with a CB₁ antagonist. From these studies, it seems that cannabinoids regulate the tissue response to inflammation in the colon. It is possible that this regulation occurs on two levels: the first, involving the smooth-muscle response to pro-inflammatory mediators that affect gastrointestinal transit time; and the second, involving the direct suppression of pro-inflammatory-mediator production (FIG. 3). Further studies will be needed to determine the relative contribution of each of these possibilities.

Arthritis. Several cannabinoids have been shown to be anti-inflammatory in animal models of arthritis. The first of these is the dimethylheptyl homologue of the natural *C. sativa* plant product THC-11-oic acid¹³. This derivative, known as ajulemic acid, binds cannabinoid receptors with very low affinity. It was initially tested for its effects on the AIR-POUCH INFLAMMATORY RESPONSE in mice and on adjuvant-induced arthritis in rats¹³ (TABLE 3). Daily feeding of ajulemic acid suppressed leukocyte accumulation after injection of IL-1 β and TNF into the pouch. In the adjuvant-induced arthritis model, chronic administration (every third day) of ajulemic acid attenuated inflammation in the joints, compared with animals that did not receive ajulemic acid. In terms of the mechanism of action, ajulemic acid was shown in *in vitro* studies to suppress prostaglandin production to a greater extent than the anti-inflammatory drug INDOMETHACIN¹³. In other studies, ajulemic acid was shown to be more potent than common non-steroidal anti-inflammatory drugs in suppressing adjuvant-induced arthritis, with less gastrointestinal ulceration occurring¹⁰⁹, and it was shown to decrease the LPS-induced production of IL-1 β in cultures of human PBMCs or synovial-fluid monocytes¹¹⁰. So, ajulemic acid might have therapeutic potential for patients with arthritis or other chronic inflammatory diseases, and it could have fewer side-effects than conventional therapies; however, there is some controversy concerning its psychoactive potential¹¹¹.

The second non-psychoactive component of *C. sativa* that has anti-inflammatory potential is cannabidiol^{112,113}. This drug was tested in a mouse model of collagen-induced arthritis, and when administered either intraperitoneally or orally (every day), beginning at the first signs of arthritis, it effectively blocked progression of the disease. It seemed to function as an

GLUTAMATERGIC SYNAPTIC TRANSMISSION

Glutamic acid is the main excitatory transmitter in the central nervous system, where it mediates fast synaptic transmission. It is released from the terminal of a glutamatergic nerve, crosses the synaptic cleft and acts on postsynaptic receptors.

AIR-POUCH INFLAMMATORY RESPONSE

An experimental model of acute inflammation. Skin pouches are established on the backs of mice, by subcutaneous injection of air on several consecutive days. Subsequently, inflammation is induced by injection of interleukin-1 β and tumour-necrosis factor into the pouch cavity.

INDOMETHACIN

A cyclooxygenase inhibitor and thereby a non-steroidal anti-inflammatory drug.

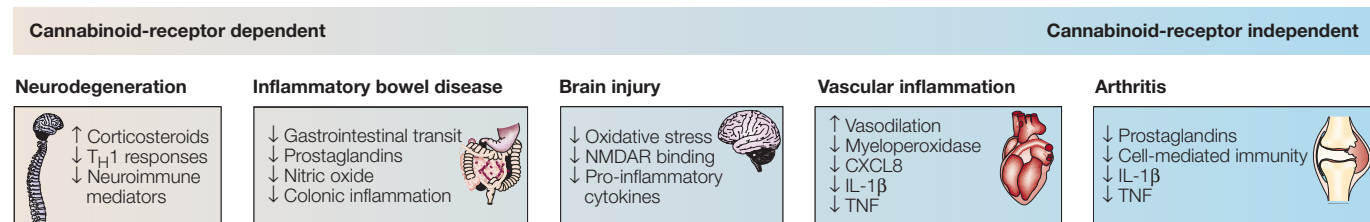


Figure 3 | Anti-inflammatory effects of cannabinoid-based drugs. Symptoms of neurodegeneration are attenuated by treatment with cannabinoid-based drugs. This process is mediated, at least in part, through binding of cannabinoid receptors. It involves an increase in corticosteroid release and a decrease in both T helper 1 (T_H1)-cell responses in the central nervous system and in neuroimmune-mediator production, including histamine and tumour-necrosis factor (TNF). Similarly, brain oedema that occurs after injury is suppressed by treatment with cannabinoid-based drugs. This process is also mediated, at least in part, through binding of cannabinoid receptors. It involves a decrease in oxidative stress, NMDA (*N*-methyl-D-aspartate) receptor (NMDAR) binding and cytokine production, including TNF. The pathophysiology of inflammatory bowel disease is also suppressed by cannabinoids. This involves binding to cannabinoid receptors, and there is a concomitant decrease in gastrointestinal-tract faecal transit, prostaglandin and nitric-oxide production, and colonic-tissue inflammation. The inflammation that is associated with arthritis is suppressed by cannabinoids. This occurs through cannabinoid-receptor-independent mechanisms, which involve a decrease in prostaglandin production and cell-mediated immunity, together with a decrease in cytokine production, including interleukin-1β (IL-1β) and TNF. Finally, vascular inflammatory disease might be partly regulated through the binding of cannabinoids to cannabinoid receptors and through vasodilation effects. This occurs together with a decrease in local pro-inflammatory-mediator production, including myeloperoxidase, CXC-chemokine ligand 8 (CXCL8), IL-1β and TNF, through cannabinoid-receptor-dependent or -independent mechanisms.

immunosuppressant, because cells from the draining lymph node of treated mice showed reduced cell-mediated immune functions and IFN-γ production in response to stimulation with antigen. Furthermore, treatment with cannabidiol blocked the LPS-induced increase in serum TNF and other immune responses. In a subsequent study, similar results were obtained at lower doses using a more potent dimethylheptyl derivative of cannabidiol¹¹⁴. These data indicate that plant-derived cannabinoids and their synthetic derivatives are anti-inflammatory and immunosuppressive; their mechanisms of action are independent of cannabinoid receptors and are mediated, in part, by suppression of pro-inflammatory-cytokine production by lymphocytes and macrophages (FIG. 3).

Vascular inflammation. Many of the features of cardiovascular disease and atherosclerosis include the elements and mechanisms of the inflammatory cascade, and this contributes to thrombosis and tissue destruction. This is also true of the pathophysiology of septic shock, in which pro-inflammatory mediators are released into the blood in excess, leading to thrombosis, vasodilation, capillary leakage and organ-system failure. Recent evidence indicates that cannabinoids might influence vascular inflammatory disease in several ways. One way is as a mediator of vasodilation, particularly LPS-induced hypotension^{4,115}. Levels of the endocannabinoids AEA and 2-AG were found to increase in the sera of patients with endotoxic shock¹¹⁶, and CB₁ antagonists were shown to prevent LPS-induced hypotension in animals, indicating that endocannabinoids and CB₁ might function as an endogenous system that promotes LPS-induced hypotension¹¹⁵ (TABLE 3). In addition to this effect of CB₁, there is also evidence of a second way in which vascular inflammatory disease might be influenced, because CB₂ is involved in vascular changes during inflammation. Myocardial ischaemia–reperfusion

injury in mice, as measured by infarct size, was considerably reduced by pretreatment with WIN55,212-2, and the protective effect was attenuated by a CB₂ antagonist but not a CB₁ antagonist⁵⁸. Treatment with WIN55,212-2 also reduced the levels of myeloperoxidase, IL-1β and CXCL8 in injured tissue, indicating that it suppresses the mobilization of pro-inflammatory mediators. The authors of this study speculated that WIN55,212-2 might have exerted its effect by binding CB₂ at the cell surface of macrophages in the injured tissue⁵⁸. There is also a third mechanism in that cannabinoid effects that are independent of both CB₁ and CB₂ have been shown to attenuate septic shock¹¹⁷. The non-psychoactive cannabinoid HU-211, when given as single pretreatment dose, reduced mouse and rat lethality in response to a challenge with LPS. In addition, similar to effects in models of closed head injury, HU-211 markedly suppressed TNF production, which might explain the attenuation of septic shock. So, it seems that cannabinoids have both a hypotensive effect that is CB₁ mediated and an anti-inflammatory effect that is either CB₂ mediated or independent of cannabinoid receptors (FIG. 3). This complexity provides both challenges and opportunities in the management of cardiovascular diseases.

Conclusions

Several general principles emerge from the studies that are discussed in this Review. The first is that endocannabinoids are expressed by immune cells and that cannabinoid receptors at the surface of immune cells are activated after infection or immune stimulation. The consequences of this for the immune response are not fully understood, but regulation of cellular chemotaxis seems to be involved. Furthermore, because cannabinoid receptors are G-protein-coupled receptors (similar to receptors for chemokines and lipid mediators), their ligation during immune stimulation probably leads to regulation of gene products that are

required for immune-cell function. The second general principle is that the main immune targets of cannabinoid-based drugs involve the suppression of cytokines and cell-mediated immunity, through cannabinoid-receptor-dependent and -independent mechanisms. The cannabinoid-receptor-dependent mechanisms most probably involve G-protein signalling and the regulation of numerous cytokine genes. Regarding cannabinoid-receptor-independent mechanisms, in the case of endocannabinoids, their effects could be mediated through vanilloid receptors, which are known to bind these compounds¹¹⁸, or they might result from the binding of as-yet-uncharacterized receptors¹¹⁹. Cannabinoid-receptor-independent mechanisms might also involve effects on lipid-raft structure and function¹²⁰,

which are known to be important for immune-cell function¹²¹. The third general principle involves the suppression, by marijuana-based drugs, of the chronic inflammatory response and the subsequent attenuation of disease processes and symptoms. These anti-inflammatory effects are undoubtedly, in part, associated with the ability of these drugs to suppress the expression of cytokines, as well as other endogenous pro-inflammatory mediators. In addition, these drugs might also function by increasing the production of anti-inflammatory mediators. Further elucidation of the effect of marijuana-based drugs on pro-inflammatory and anti-inflammatory mechanisms will provide the basis for the formulation of more effective drugs for the management of chronic inflammatory diseases.

- Tomida, I., Pertwee, R. G. & Azuara-Blanco, A. Cannabinoids and glaucoma. *Br. J. Ophthalmol.* **88**, 708–713 (2004).
- Guzman, M. Cannabinoids: potential anticancer agents. *Nature Rev. Cancer* **3**, 745–755 (2003).
- Kunos, G. & Pacher, P. Cannabinoids cool the intestine. *Nature Med.* **10**, 678–679 (2004).
- Mendizabal, V. E. & Adler-Graschinsky, E. Cannabinoid system as a potential target for drug development in the treatment of cardiovascular disease. *Curr. Vasc. Pharmacol.* **1**, 301–313 (2003).
- Mechoulam, R., Panikashvili, D. & Shohami, E. Cannabinoids and brain injury: therapeutic implications. *Trends Mol. Med.* **8**, 58–61 (2002).
- Baker, D. & Pryce, G. The therapeutic potential of cannabis in multiple sclerosis. *Expert Opin. Investig. Drugs* **12**, 561–567 (2003).
- Baker, D., Pryce, G., Giovannoni, G. & Thompson, A. J. The therapeutic potential of cannabis. *Lancet Neurol.* **2**, 291–298 (2003).
- Di Marzo, V., Bifulco, M. & De Petrocellis, L. The endocannabinoid system and its therapeutic exploitation. *Nature Rev. Drug Discov.* **3**, 771–784 (2004).
- Black, S. C. Cannabinoid receptor antagonists and obesity. *Curr. Opin. Investig. Drugs* **5**, 389–394 (2004).
- Gaoni, Y. & Mechoulam, R. Isolation, structure, and partial synthesis of an active constituent of hashish. *J. Am. Chem. Soc.* **86**, 1646–1647 (1964).
This is a seminal paper that describes the isolation and synthesis of THC.
- Howlett, A. C. *et al.* International Union of Pharmacology. XXVII. Classification of cannabinoid receptors. *Pharmacol. Rev.* **54**, 161–202 (2002).
For those wanting more on the chemistry and biology of the cannabinoid system, this is a recent and authoritative review.
- Rhee, M. H. *et al.* Cannabinoid derivatives: binding to cannabinoid receptors and inhibition of adenylyl cyclase. *J. Med. Chem.* **40**, 3228–3233 (1997).
- Zurier, R. B. *et al.* Dimethylheptyl-THC-11 oic acid. A nonpsychoactive antiinflammatory agent with a cannabinoid template structure. *Arthritis Rheum.* **41**, 163–170 (1998).
- Burstein, S. H. The cannabinoid acids: nonpsychoactive derivatives with therapeutic potential. *Pharmacol. Ther.* **82**, 87–96 (1999).
- Liu, J., Li, H., Burstein, S. H., Zurier, R. B. & Chen, J. D. Activation and binding of peroxisome proliferator-activated receptor γ by synthetic cannabinoid ajulemic acid. *Mol. Pharmacol.* **63**, 983–992 (2003).
Cannabinoid-based drugs might function through mechanisms other than binding cannabinoid receptors, such as through binding PPAR- γ , as described in this paper.
- Piomelli, D. The molecular logic of endocannabinoid signalling. *Nature Rev. Neurosci.* **4**, 873–884 (2003).
Cannabinoids are best known for their psychoactive effects on the brain. This Review, together with references 28 and 29, discusses these mechanisms in detail.
- Sugiura, T., Kobayashi, Y., Oka, S. & Waku, K. Biosynthesis and degradation of anandamide and 2-arachidonoylglycerol and their possible physiological significance. *Prostaglandins Leukot. Essent. Fatty Acids* **66**, 173–192 (2002).
- Devane, W. A. *et al.* Isolation and structure of a brain constituent that binds to the cannabinoid receptor. *Science* **258**, 1946–1949 (1992).
An important breakthrough in the field was the discovery of an endocannabinoid. This paper describes the isolation of the first endocannabinoid to be found, AEA (previously known as anandamide).
- Zygmunt, P. M. *et al.* Vanilloid receptors on sensory nerves mediate the vasodilator action of anandamide. *Nature* **400**, 452–457 (1999).
- Mechoulam, R. *et al.* Identification of an endogenous 2-monoglyceride, present in canine gut, that binds to cannabinoid receptors. *Biochem. Pharm.* **50**, 83–90 (1995).
- Hanus, L. *et al.* 2-Arachidonoyl glyceryl ether, an endogenous agonist of the cannabinoid CB₁ receptor. *Proc. Natl Acad. Sci. USA* **98**, 3662–3665 (2001).
- Matsuda, L. A., Lolait, S. J., Brownstein, M. J., Young, A. C. & Bonner, T. I. Structure of cannabinoid receptor and functional expression of the cloned cDNA. *Nature* **346**, 561–564 (1990).
- Munro, S., Thomas, K. L. & Abu-Shaar, M. Molecular characterization of a peripheral receptor for cannabinoids. *Nature* **365**, 61–65 (1993).
References 20–23 describe the isolation of endocannabinoids that were discovered subsequently to AEA and the isolation of the cannabinoid receptors, CB₁ and CB₂.
- McAllister, S. D. & Glass, M. CB₁ and CB₂ receptor-mediated signalling: a focus on endocannabinoids. *Prostaglandins Leukot. Essent. Fatty Acids* **66**, 161–171 (2002).
- Klein, T. W. *et al.* The cannabinoid system and immune modulation. *J. Leukoc. Biol.* **74**, 486–496 (2003).
- Klein, T., Newton, C. & Friedman, H. Cannabinoid receptors and immunity. *Immunol. Today* **19**, 373–381 (1998).
- Berdyshev, E. V. Cannabinoid receptors and the regulation of immune response. *Chem. Phys. Lipids* **108**, 169–190 (2000).
- Schlicker, E. & Kathmann, M. Modulation of transmitter release via presynaptic cannabinoid receptors. *Trends Pharmacol. Sci.* **22**, 565–572 (2001).
- Christie, M. J. & Vaughan, C. W. Cannabinoids act backwards. *Nature* **410**, 527–530 (2001).
- Akira, S. & Takeda, K. Toll-like receptor signalling. *Nature Rev. Immunol.* **4**, 499–511 (2004).
- Noveri, M. C., Erb-Downward, J. R. & Huffnagle, G. B. Production of eicosanoids and other oxylipins by pathogenic eukaryotic microbes. *Clin. Microbiol. Rev.* **16**, 517–533 (2003).
- DiMarzo, V., DePetrocellis, L., Sepe, N. & Buono, A. Biosynthesis of anandamide and related acylethanolamides in mouse J774 macrophages and N18 neuroblastoma cells. *Biochem. J.* **316**, 977–984 (1996).
- DiMarzo, V. *et al.* Biosynthesis and inactivation of the endocannabinoid 2-arachidonoylglycerol in circulating and tumoral macrophages. *Eur. J. Biochem.* **264**, 258–267 (1999).
- Maccarrone, M., Bari, M., Battista, N. & Finazzi-Agro, A. Endocannabinoid degradation, endotoxic shock and inflammation. *Curr. Drug Targets Inflamm. Allergy* **1**, 53–63 (2002).
- Matias, I. *et al.* Presence and regulation of the endocannabinoid system in human dendritic cells. *Eur. J. Biochem.* **269**, 3771–3778 (2002).
- Maccarrone, M. *et al.* Lipopolysaccharide downregulates fatty acid amide hydrolase expression and increases anandamide levels in human peripheral lymphocytes. *Arch. Biochem. Biophys.* **393**, 321–328 (2001).
References 32–36 discuss the metabolism of endocannabinoids in immune cells.
- Wyss-Coray, T. & Mucke, L. Inflammation in neurodegenerative disease — a double-edged sword. *Neuron* **35**, 419–432 (2002).
- Moser, B., Wolf, M., Walz, A. & Loetscher, P. Chemokines: multiple levels of leukocyte migration control. *Trends Immunol.* **25**, 75–84 (2004).
- Oka, S. *et al.* 2-Arachidonoylglycerol, an endogenous cannabinoid receptor ligand, induces the migration of EoL-1 human eosinophilic leukemia cells and human peripheral blood eosinophils. *J. Leukoc. Biol.* **76**, 1002–1009 (2004).
- Rayman, N. *et al.* Distinct expression profiles of the peripheral cannabinoid receptor in lymphoid tissues depending on receptor activation status. *J. Immunol.* **172**, 2111–2117 (2004).
- Maestroni, G. J. The endogenous cannabinoid 2-arachidonoyl glycerol as *in vivo* chemoattractant for dendritic cells and adjuvant for T_H1 response to a soluble protein. *FASEB J.* **18**, 1914–1916 (2004).
- Alberich Jorda, M. *et al.* The peripheral cannabinoid receptor CB₂, frequently expressed on AML blasts, either induces a neutrophilic differentiation block or confers abnormal migration properties in a ligand-dependent manner. *Blood* **104**, 526–534 (2004).
The many putative roles of cannabinoid receptors in immune-cell biology remain undefined. This paper indicates that these receptors might have a role in leukaemia.
- Szabo, I. *et al.* Heterologous desensitization of opioid receptors by chemokines inhibits chemotaxis and enhances the perception of pain. *Proc. Natl Acad. Sci. USA* **99**, 10276–10281 (2002).
- Lee, S. F., Newton, C., Widen, R., Friedman, H. & Klein, T. W. Differential expression of cannabinoid CB₂ receptor mRNA in mouse immune cell subpopulations and following B cell stimulation. *Eur. J. Pharmacol.* **423**, 235–241 (2001).
- Daaka, Y., Friedman, H. & Klein, T. W. Cannabinoid receptor proteins are increased in Jurkat, human T-cell line after mitogen activation. *J. Pharmacol. Exp. Ther.* **276**, 776–783 (1996).
This report was among the first to show that cannabinoid-receptor expression changes with variations in immune-cell activation.
- Noe, S. N., Newton, C., Widen, R., Friedman, H. & Klein, T. W. Anti-CD40, anti-CD3, and IL-2 stimulation induce contrasting changes in CB₂ mRNA expression in mouse splenocytes. *J. Neuroimmunol.* **110**, 161–167 (2000).
- Gardner, B. *et al.* Autocrine and paracrine regulation of lymphocyte CB₂ receptor expression by TGF- β . *Biochem. Biophys. Res. Commun.* **290**, 91–96 (2002).
- Klein, T. W., Newton, C., Zhu, W., Daaka, Y. & Friedman, H. Δ^9 -Tetrahydrocannabinol, cytokines and immunity to *Legionella pneumophila*. *Proc. Soc. Exp. Biol. Med.* **209**, 205–212 (1995).
- Carlisle, S., Marciano-Cabral, F., Staab, A., Ludwick, C. & Cabral, G. Differential expression of the CB₂ cannabinoid receptor by rodent macrophages and macrophage-like cells in relation to cell activation. *Int. Immunopharmacol.* **2**, 69–82 (2002).

50. Walter, L. *et al.* Nonpsychotropic cannabinoid receptors regulate microglial cell migration. *J. Neurosci.* **23**, 1398–1405 (2003).
Although CB₂ has been reported to be expressed in the brain, this was the first report to indicate that it might be expressed by brain microglial cells.
51. Bromley, S. K. *et al.* The immunological synapse. *Annu. Rev. Immunol.* **19**, 375–396 (2001).
52. Blanchard, D. K., Newton, C., Klein, T. W., Stewart, W. E. & Friedman, H. *In vitro* and *in vivo* suppressive effects of Δ⁹-tetrahydrocannabinol on interferon production by murine spleen cells. *Int. J. Immunopharmacol.* **8**, 819–824 (1986).
53. Cabral, G. A., Lockmuller, J. C. & Mishkin, E. M. Δ⁹-Tetrahydrocannabinol decreases α/β interferon response to herpes simplex virus type 2 in the B6C3F1 mouse. *Proc. Soc. Exp. Biol. Med.* **181**, 305–311 (1986).
54. Klein, T. W., Lane, B., Newton, C. A. & Friedman, H. The cannabinoid system and cytokine network. *Proc. Soc. Exp. Biol. Med.* **225**, 1–8 (2000).
55. Smith, S. R., Terminelli, C. & Denhardt, G. Effects of cannabinoid receptor agonist and antagonist ligands on production of inflammatory cytokines and anti-inflammatory interleukin-10 in endotoxemic mice. *J. Pharmacol. Exp. Ther.* **293**, 136–150 (2000).
56. Shohami, E., Gallily, R., Mechoulam, R., Bass, R. & Ben-Hur, T. Cytokine production in the brain following closed head injury: dexamnabinol (HU-211) is a novel TNF-α inhibitor and an effective neuroprotectant. *J. Neuroimmunol.* **72**, 169–177 (1997).
Using an animal model of closed head injury, this paper shows that non-psychoactive cannabinoids can suppress the production of pro-inflammatory cytokines.
57. Doble, A. The role of excitotoxicity in neurodegenerative disease: implications for therapy. *Pharmacol. Ther.* **81**, 163–221 (1999).
58. Di Filippo, C., Rossi, F., Rossi, S. & D'Amico, M. Cannabinoid CB₂ receptor activation reduces mouse myocardial ischemia-reperfusion injury: involvement of cytokine/chemokines and PMN. *J. Leukoc. Biol.* **75**, 453–459 (2004).
59. Baldwin, G. C. *et al.* Marijuana and cocaine impair alveolar macrophage function and cytokine production. *Am. J. Respir. Crit. Care Med.* **156**, 1606–1613 (1997).
This is one of the few reports that shows a direct suppressive effect of marijuana smoking on immune-cell function.
60. Klein, T. W., Newton, C., Widen, R. & Friedman, H. Δ⁹-Tetrahydrocannabinol injection induces cytokine-mediated mortality of mice infected with *Legionella pneumophila*. *J. Pharmacol. Exp. Ther.* **267**, 635–640 (1993).
61. Zhu, W., Newton, C., Daaka, Y., Friedman, H. & Klein, T. W. Δ⁹-Tetrahydrocannabinol enhances the secretion of interleukin 1 from endotoxin-stimulated macrophages. *J. Pharmacol. Exp. Ther.* **270**, 1334–1339 (1994).
62. Smith, S. R., Terminelli, C. & Denhardt, G. Modulation of cytokine responses in *Corynebacterium parvum*-primed endotoxemic mice by centrally administered cannabinoid ligands. *Eur. J. Pharmacol.* **425**, 73–83 (2001).
63. Derocq, J. M. *et al.* Genomic and functional changes induced by the activation of the peripheral cannabinoid receptor CB₂ in the promyelocytic cells HL-60. Possible involvement of the CB₂ receptor in cell differentiation. *J. Biol. Chem.* **275**, 15621–15628 (2000).
64. Kishimoto, S. *et al.* 2-Arachidonoylglycerol, an endogenous cannabinoid receptor ligand, induces accelerated production of chemokines in HL-60 cells. *J. Biochem. (Tokyo)* **135**, 517–524 (2004).
65. Pryce, G. *et al.* Cannabinoids inhibit neurodegeneration in models of multiple sclerosis. *Brain* **126**, 2191–2202 (2003).
66. Ni, X. *et al.* WIN 55212-2, a cannabinoid receptor agonist, attenuates leukocyte/endothelial interactions in an experimental autoimmune encephalomyelitis model. *Mult. Scler.* **10**, 158–164 (2004).
67. Kawai, T. *et al.* Selective diapedesis of T_H1 cells induced by endothelial cell RANTES. *J. Immunol.* **163**, 3269–3278 (1999).
68. Savinov, A. Y., Wong, F. S. & Chervonsky, A. V. IFN-γ affects homing of diabetogenic T cells. *J. Immunol.* **167**, 6637–6643 (2001).
69. Cabral, G. & Dove Pettit, D. Drugs and immunity: cannabinoids and their role in decreased resistance to infectious diseases. *J. Neuroimmunol.* **83**, 116–123 (1998).
70. Newton, C. A., Klein, T. W. & Friedman, H. Secondary immunity to *Legionella pneumophila* and T_H1 activity are suppressed by Δ⁹-tetrahydrocannabinol injection. *Infect. Immun.* **62**, 4015–4020 (1994).
This was the first report to show that cannabinoids bias the immune response away from T_H1-cell responses.
71. Klein, T. W., Newton, C. A., Nakachi, N. & Friedman, H. Δ⁹-Tetrahydrocannabinol treatment suppresses immunity and early IFN_γ, IL-12, and IL-12 receptor β2 responses to *Legionella pneumophila* infection. *J. Immunol.* **164**, 6461–6466 (2000).
72. Braun, M. C. & Kelsall, B. L. Regulation of interleukin-12 production by G-protein-coupled receptors. *Microbes Infect.* **3**, 99–107 (2001).
73. Zhu, L. X. *et al.* Δ⁹-Tetrahydrocannabinol inhibits antitumor immunity by a CB₂ receptor-mediated, cytokine-dependent pathway. *J. Immunol.* **165**, 373–380 (2000).
74. Piccinillo, C. A. & Thornton, A. M. Cornerstone of peripheral tolerance: naturally occurring CD4⁺CD25⁺ regulatory T cells. *Trends Immunol.* **25**, 374–380 (2004).
75. Weiner, H. L. Induction and mechanism of action of transforming growth factor-β-secreting T_H3 regulatory cells. *Immunol. Rev.* **182**, 207–214 (2001).
76. Yuan, M. *et al.* Δ⁹-Tetrahydrocannabinol regulates T_H1/T_H2 cytokine balance in activated human T cells. *J. Neuroimmunol.* **133**, 124–131 (2002).
77. Pacifici, R. *et al.* Modulation of the immune system in cannabis users. *JAMA* **289**, 1929–1931 (2003).
This report provided the first evidence from a human study to support the findings in animals that indicate that marijuana-derived cannabinoids bias T_H1-cell responses.
78. Croxford, J. L. & Miller, S. D. Immunoregulation of a viral model of multiple sclerosis using the synthetic cannabinoid R(+)-WIN55,212. *J. Clin. Invest.* **111**, 1231–1240 (2003).
79. Arevalo-Martin, A., Vela, J. M., Molina-Holgado, E., Borrell, J. & Guaza, C. Therapeutic action of cannabinoids in a murine model of multiple sclerosis. *J. Neurosci.* **23**, 2511–2516 (2003).
80. Das, J. *et al.* A critical role for NF-κB in *Gata3* expression and T_H2 differentiation in allergic airway inflammation. *Nature Immunol.* **2**, 45–50 (2001).
81. Klein, T. W. *et al.* Cannabinoid receptors and T helper biasing. *J. Neuroimmunol.* **147**, 91–94 (2004).
82. Herring, A. C. & Kaminski, N. E. Cannabinoid-mediated inhibition of nuclear factor-κB, cAMP response element-binding protein, and interleukin-2 secretion by activated thymocytes. *J. Pharmacol. Exp. Ther.* **291**, 1156–1163 (1999).
83. Foulds, J., Burke, M., Steinberg, M., Williams, J. M. & Ziedonis, D. M. Advances in pharmacotherapy for tobacco dependence. *Expert Opin. Emerg. Drugs* **9**, 39–53 (2004).
84. Howlett, A. C. *et al.* Cannabinoid physiology and pharmacology: 30 years of progress. *Neuropharmacology* **47**, 345–358 (2004).
85. Baker, D. *et al.* Cannabinoids control spasticity and tremor in a multiple sclerosis model. *Nature* **404**, 84–87 (2000).
The central effects of cannabinoids that bind CB₂ might have a role in controlling spasticity and tremor in individuals with multiple sclerosis, as shown in animals with chronic, relapsing EAE.
86. De Petrocellis, L. *et al.* The endogenous cannabinoid anandamide inhibits human breast cancer cell proliferation. *Proc. Natl Acad. Sci. USA* **95**, 8375–8380 (1998).
87. Galve-Roperh, I. *et al.* Anti-tumoral action of cannabinoids: involvement of sustained ceramide accumulation and extracellular signal-regulated kinase activation. *Nature Med.* **6**, 313–319 (2000).
88. Bifulco, M. *et al.* A new strategy to block tumor growth by inhibiting endocannabinoid inactivation. *FASEB J.* **18**, 1606–1608 (2004).
In this paper, a novel mechanism is reported for endocannabinoids in the inhibition of tumour growth.
89. Lyman, W. D., Sonett, J. R., Brosnan, C. F., Elkin, R. & Bornstein, M. B. Δ⁹-Tetrahydrocannabinol: a novel treatment for experimental autoimmune encephalomyelitis. *J. Neuroimmunol.* **23**, 73–81 (1989).
90. Wirguin, I. *et al.* Suppression of experimental autoimmune encephalomyelitis by cannabinoids. *Immunopharmacology* **28**, 209–214 (1994).
91. Newton, C. A. *et al.* The THC-induced suppression of T_H1 polarization in response to *Legionella pneumophila* infection is not mediated by increases in corticosterone and PGE₂. *J. Leukoc. Biol.* **76**, 854–861 (2004).
92. Kingery, W. S. A critical review of controlled clinical trials for peripheral neuropathic pain and complex regional pain syndromes. *Pain* **73**, 123–139 (1997).
93. Calignano, A., La Rana, G., Giuffrida, A. & Piomelli, D. Control of pain initiation by endogenous cannabinoids. *Nature* **394**, 277–281 (1998).
94. Ibrahim, M. M. *et al.* Activation of CB₂ cannabinoid receptors by AM1241 inhibits experimental neuropathic pain: pain inhibition by receptors not present in the CNS. *Proc. Natl Acad. Sci. USA* **100**, 10529–10533 (2003).
This study shows that cannabinoids might inhibit pain by binding CB₂, as well as CB₁.
95. Svendsen, K. B., Jensen, T. S. & Bach, F. W. Does the cannabinoid dronabinol reduce central pain in multiple sclerosis? Randomised double blind placebo controlled crossover trial. *BMJ* **329**, 253 (2004).
96. Baker, D. *et al.* Endocannabinoids control spasticity in a multiple sclerosis model. *FASEB J.* **15**, 300–302 (2001).
97. Zajicek, J. *et al.* Cannabinoids for treatment of spasticity and other symptoms related to multiple sclerosis (CAMS study): multicentre randomised placebo-controlled trial. *Lancet* **362**, 1517–1526 (2003).
98. Vaney, C. *et al.* Efficacy, safety and tolerability of an orally administered cannabis extract in the treatment of spasticity in patients with multiple sclerosis: a randomized, double-blind, placebo-controlled, crossover study. *Mult. Scler.* **10**, 417–424 (2004).
99. Wade, D. T., Makela, P., Robson, P., House, H. & Bateman, C. Do cannabis-based medicinal extracts have general or specific effects on symptoms in multiple sclerosis? A double-blind, randomized, placebo-controlled study on 160 patients. *Mult. Scler.* **10**, 434–441 (2004).
100. Panikashvili, D. *et al.* An endogenous cannabinoid (2-AG) is neuroprotective after brain injury. *Nature* **413**, 527–531 (2001).
101. Knoller, N. *et al.* Dexamnabinol (HU-211) in the treatment of severe closed head injury: a randomized, placebo-controlled, Phase II clinical trial. *Crit. Care Med.* **30**, 548–554 (2002).
102. Pharms Corporation. Dexamnabinol did not demonstrate efficacy [online], <http://www.pharmscorp.com/news/pr/pr122004.html> (2004).
103. Molina-Holgado, F. *et al.* Endogenous interleukin-1 receptor antagonist mediates anti-inflammatory and neuroprotective actions of cannabinoids in neurons and glia. *J. Neurosci.* **23**, 6470–6474 (2003).
104. Manara, L. *et al.* Functional assessment of neuronal cannabinoid receptors in the muscular layers of human ileum and colon. *Dig. Liver Dis.* **34**, 262–269 (2002).
105. Izzo, A. A. *et al.* Cannabinoid CB₂-receptor mediated regulation of gastrointestinal motility in mice in a model of intestinal inflammation. *Br. J. Pharmacol.* **134**, 563–570 (2001).
106. Pinto, L. *et al.* Endocannabinoids as physiological regulators of colonic propulsion in mice. *Gastroenterology* **123**, 227–234 (2002).
107. Mathison, R., Ho, W., Pittman, Q. J., Davison, J. S. & Sharkey, K. A. Effects of cannabinoid receptor-2 activation on accelerated gastrointestinal transit in lipopolysaccharide-treated rats. *Br. J. Pharmacol.* **142**, 1247–1254 (2004).
108. Massa, F. *et al.* The endogenous cannabinoid system protects against colonic inflammation. *J. Clin. Invest.* **113**, 1202–1209 (2004).
The endocannabinoid system might control and regulate intestinal inflammatory responses, as reported in this paper.
109. Dajani, E. Z. *et al.* 1',1'-Dimethylheptyl-Δ⁸-tetrahydrocannabinol-11-oic acid: a novel, orally effective cannabinoid with analgesic and anti-inflammatory properties. *J. Pharmacol. Exp. Ther.* **291**, 31–38 (1999).
110. Zurier, R. B., Rossetti, R. G., Burstein, S. H. & Biding, B. Suppression of human monocyte interleukin-1β production by ajulemic acid, a nonpsychoactive cannabinoid. *Biochem. Pharmacol.* **65**, 649–655 (2003).
111. Burstein, S. & Zurier, R. B. Pain reduction and lack of psychotropic effects with ajulemic acid. *Arthritis Rheum.* **50**, 4078–4079 (2004).
112. Malfait, A. M. *et al.* The nonpsychoactive cannabis constituent cannabidiol is an oral anti-arthritic therapeutic in murine collagen-induced arthritis. *Proc. Natl Acad. Sci. USA* **97**, 9561–9566 (2000).
113. Gallily, R. *et al.* γ-Irradiation enhances apoptosis induced by cannabidiol, a non-psychoactive cannabinoid, in cultured HL-60 myeloblastic leukemia cells. *Leuk. Lymphoma* **44**, 1767–1773 (2003).
114. Sumariwalla, P. F. *et al.* A novel synthetic, nonpsychoactive cannabinoid acid (HU-320) with antiinflammatory properties in murine collagen-induced arthritis. *Arthritis Rheum.* **50**, 985–998 (2004).
References 112–114 use animal models to show the anti-inflammatory potential of non-psychoactive cannabinoids.
115. Varga, K., Wagner, J. A., Bridgen, D. T. & Kunos, G. Platelet- and macrophage-derived endogenous cannabinoids are involved in endotoxin-induced hypotension. *FASEB J.* **12**, 1035–1044 (1998).
116. Wang, Y. *et al.* Simultaneous measurement of anandamide and 2-arachidonoylglycerol by polymyxin B-selective adsorption and subsequent high-performance liquid chromatography analysis: increase in endogenous cannabinoids in the sera of patients with endotoxemic shock. *Anal. Biochem.* **294**, 73–82 (2001).

117. Gallily, R. *et al.* Protection against septic shock and suppression of tumor necrosis factor α and nitric oxide production by dexanabinol (HU-211), a nonpsychotropic cannabinoid. *J. Pharmacol. Exp. Ther.* **283**, 918–924 (1997).
118. Veldhuis, W. B. *et al.* Neuroprotection by the endogenous cannabinoid anandamide and arvanil against *in vivo* excitotoxicity in the rat: role of vanilloid receptors and lipoxygenases. *J. Neurosci.* **23**, 4127–4133 (2003).
119. Breivogel, C. S., Griffin, G., Di Marzo, V. & Martin, B. R. Evidence for a new G protein-coupled cannabinoid receptor in mouse brain. *Mol. Pharmacol.* **60**, 155–163 (2001).
This paper indicates that there might be more than two cannabinoid receptors.
120. Biswas, K. K. *et al.* Membrane cholesterol but not putative receptors mediates anandamide-induced hepatocyte apoptosis. *Hepatology* **38**, 1167–1177 (2003).
This paper shows that lipid rafts might be affected by cannabinoid-based drugs.
121. Vogt, A. B., Spindeldreher, S. & Kropshofer, H. Clustering of MHC-peptide complexes prior to their engagement in the immunological synapse: lipid raft and tetraspan microdomains. *Immunol. Rev.* **189**, 136–151 (2002).
122. Fischer-Stenger, K., Dove Pettit, D. A. & Cabral, G. A. Δ^9 -Tetrahydrocannabinol inhibition of tumor necrosis factor- α : suppression of post-translational events. *J. Pharm. Exp. Ther.* **267**, 1558–1565 (1993).
123. Chang, Y. H., Lee, S. T. & Lin, W. W. Effects of cannabinoids on LPS-stimulated inflammatory mediator release from macrophages: involvement of eicosanoids. *J. Cell. Biochem.* **81**, 715–723 (2001).
124. Facchinetti, F., Del Giudice, E., Furegato, S., Passarotto, M. & Leon, A. Cannabinoids ablate release of TNF α in rat microglial cells stimulated with lipopolysaccharide. *Glia* **41**, 161–168 (2003).

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Competing interests statement

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